Resource Summary Report

Generated by RRID on Aug 2, 2026

Monoclonal Anti-alpha-Actinin (Sarcomeric) antibody produced in mouse

RRID:AB_476766
Type: Antibody

Proper Citation

Sigma-Aldrich Cat# A7811, RRID:AB_476766

Antibody Information

URL: http://antibodyregistry.org/AB_476766

Proper Citation: Sigma-Aldrich Cat# A7811, RRID:AB_476766

Target Antigen: alpha-Actinin (Sarcomeric)

Host Organism: mouse

Clonality: monoclonal

Comments: Applications: ELISA, dot blot, immunoblot, immunohistochemistry, immunocytochemistry.

Antibody Name: Monoclonal Anti-alpha-Actinin (Sarcomeric) antibody produced in mouse

Description: This monoclonal targets alpha-Actinin (Sarcomeric)

Target Organism: chicken, feline, rat, hamster, porcine, snake, canine, goat, reptile, pig, mouse, frog, fish, rabbit, bovine, zebrafish, human, lizard, sheep

Clone ID: EA-53

Defining Citation: PMID:20235162

Antibody ID: AB_476766

Vendor: Sigma-Aldrich

Catalog Number: A7811

Record Creation Time: 20250820T000546+0000

Record Last Update: 20250820T072145+0000

Ratings and Alerts

No rating or validation information has been found for Monoclonal Anti-alpha-Actinin (Sarcomeric) antibody produced in mouse.

No alerts have been found for Monoclonal Anti-alpha-Actinin (Sarcomeric) antibody produced in mouse.

Data and Source Information

Source: Antibody Registry

Usage and Citation Metrics

We found 147 mentions in open access literature.

Listed below are recent publications. The full list is available at RRID .

Liu Y, et al. (2026) Human PSC-derived organoids model sympathetic ganglion development and its functional crosstalk with the heart. Cell stem cell, 33(1), 29.

Mohammadi N, et al. (2025) Sympathetic neurons can modify the intrinsic structural and functional properties of human pluripotent stem cell-derived cardiomyocytes. The Journal of physiology, 603(7), 2089.

Eggertsen TG, et al. (2025) Logic-based machine learning predicts how escitalopram attenuates cardiomyocyte hypertrophy. Proceedings of the National Academy of Sciences of the United States of America, 122(10), e2420499122.

Wang Y, et al. (2025) Injectable hydrogel with miR-222-engineered extracellular vesicles ameliorates myocardial ischemic reperfusion injury via mechanotransduction. Cell reports. Medicine, 6(3), 101987.

Kaltenecker D, et al. (2025) Functional liver genomics identifies hepatokines promoting wasting in cancer cachexia. Cell, 188(17), 4549.

Wang W, et al. (2025) Zbtb16 determines the fate plasticity of cardiovascular progenitors through IGF2BP3-mediated mRNA stabilization. Cell reports, 44(8), 116127.

Raabe J, et al. (2025) Generation of a biallelic NRAP-knockout mutant from a human iPSC line. Stem cell research, 88, 103829.

Lin S, et al. (2025) Rbpms2 prevents major cardiac defects in cardiomyocyte-specific Rbpms-deficient mice. Developmental cell.

Zhou F, et al. (2025) Stabilisation of PRCP by deubiquitinase-targeting chimera (DUBTAC) to replenish autophagy for ameliorating pathological cardiac hypertrophy. British journal of pharmacology.

Tanaka S, et al. (2025) Fatty acid metabolism suppresses neonatal cardiomyocyte proliferation by increasing PDK4 and HMGCS2 expression through PPAR??. PloS one, 20(5), e0318178.

Durdu S, et al. (2025) Chromatin-dependent motif syntax defines differentiation trajectories. Molecular cell, 85(15), 2900.

Valencia DA, et al. (2025) Human formin FHOD3-mediated actin elongation is required for sarcomere integrity in cardiomyocytes. eLife, 13.

Feeney AK, et al. (2025) Enhancing human pluripotent stem cell differentiation to cardiomyocytes through cardiac progenitor reseeding and cryopreservation. iScience, 28(5), 112452.

Swift SK, et al. (2024) A broadly applicable method for quantifying cardiomyocyte cell division identifies proliferative events following myocardial infarction. Cell reports methods, 4(9), 100860.

Ugorets V, et al. (2024) Dynamic remodeling of septin structures fine-tunes myogenic differentiation. iScience, 27(9), 110630.

Syed Ali G, et al. (2024) Generation of a heterozygous Calsequestrin 2 F189L iPSC line (UMGi158-B) by CRISPR/Cas9 genome editing to investigate the cardiac pathophysiology of Takotsubo Syndrome and Catecholaminergic Polymorphic Ventricular Tachycardia. Stem cell research, 81, 103538.

Duboscq-Bidot L, et al. (2024) Generation of CRISPR-Cas9 edited human induced pluripotent stem cell line carrying BAG3 V468M mutation in its BAG domain. Stem cell research, 74, 103294.

Schreiber MK, et al. (2024) Generation of Pelizaeus-Merzbacher disease (PMD) mutant (PLP1-C33Y) in induced pluripotent stem cell (iPSC) by CRISPR/Cas9 genome editing. Stem cell research, 74, 103276.

Busley AV, et al. (2024) Mutation-induced LZTR1 polymerization provokes cardiac pathology in recessive Noonan syndrome. Cell reports, 43(7), 114448.

Pierre B, et al. (2024) Generation of CRISPR/Cas9 edited human induced pluripotent stem cell line carrying the heterozygous p.H695VfsX5 frameshift mutation in the exon 10 of the PKP2 gene. Stem cell research, 76, 103341.