Resource Summary Report

Generated by RRID on Aug 19, 2026

p415-GalL-Cas9-CYC1t

RRID:Addgene_43804
Type: Plasmid

Proper Citation

RRID:Addgene_43804

Plasmid Information

URL: http://www.addgene.org/43804

Proper Citation: RRID:Addgene_43804

Insert Name: Human Optimized S. pyogenes Cas9

Organism: Homo sapiens

Bacterial Resistance: Ampicillin

Defining Citation: PMID:23460208

Vector Backbone Description: Backbone Size:6670; Vector Backbone:p415; Vector Types:Yeast Expression, CRISPR; Bacterial Resistance:Ampicillin

Plasmid Name: p415-GalL-Cas9-CYC1t

Record Creation Time: 20220422T222227+0000

Record Last Update: 20260815T081510+0000

Ratings and Alerts

No rating or validation information has been found for p415-GalL-Cas9-CYC1t.

No alerts have been found for p415-GalL-Cas9-CYC1t.

Data and Source Information

Source: Addgene

Usage and Citation Metrics

We found 18 mentions in open access literature.

Listed below are recent publications. The full list is available at RRID .

Vedova CD, et al. (2025) Combining face-to-face laboratory sessions and a computer simulation effectively teaches gene editing and DNA sequencing to undergraduate genetics students. Biochemistry and molecular biology education : a bimonthly publication of the International Union of Biochemistry and Molecular Biology, 53(3), 286.

Boocock J, et al. (2025) Single-cell eQTL mapping in yeast reveals a tradeoff between growth and reproduction. eLife, 13.

Wang Y, et al. (2025) A unique mechanism of snRNP core assembly. Nature communications, 16(1), 3166.

Ruta GV, et al. (2024) Eukaryotic-driven directed evolution of Cas9 nucleases. Genome biology, 25(1), 79.

Mendoza BJ, et al. (2023) Potency of CRISPR-Cas Antifungals Is Enhanced by Cotargeting DNA Repair and Growth Regulatory Machinery at the Genetic Level. ACS infectious diseases, 9(12), 2494.

Barber JN, et al. (2021) The evolution of coexistence from competition in experimental co-cultures of Escherichia coli and Saccharomyces cerevisiae. The ISME journal, 15(3), 746.

Liu M, et al. (2021) High-Level Production of Sesquiterpene Patchoulol in Saccharomyces cerevisiae. ACS synthetic biology, 10(1), 158.

Mosbach V, et al. (2020) Resection and repair of a Cas9 double-strand break at CTG trinucleotide repeats induces local and extensive chromosomal deletions. PLoS genetics, 16(7), e1008924.

Shao Y, et al. (2019) Creating functional chromosome fusions in yeast with CRISPR-Cas9. Nature protocols, 14(8), 2521.

Gu Y, et al. (2019) Construction of a series of episomal plasmids and their application in the development of an efficient CRISPR/Cas9 system in Pichia pastoris. World journal of microbiology & biotechnology, 35(6), 79.

Standage-Beier K, et al. (2019) RNA-Guided Recombinase-Cas9 Fusion Targets Genomic DNA Deletion and Integration. The CRISPR journal, 2(4), 209.

Laughery MF, et al. (2019) R-loop formation by dCas9 is mutagenic in Saccharomyces cerevisiae. Nucleic acids research, 47(5), 2389.

Dank A, et al. (2018) CRISPR-Cas genome engineering of esterase activity in Saccharomyces cerevisiae steers aroma formation. BMC research notes, 11(1), 682.

Casini A, et al. (2018) A highly specific SpCas9 variant is identified by in vivo screening in yeast. Nature biotechnology, 36(3), 265.

Despr??s PC, et al. (2018) Double Selection Enhances the Efficiency of Target-AID and Cas9-Based Genome Editing in Yeast. G3 (Bethesda, Md.), 8(10), 3163.

Giersch RM, et al. (2017) Method for Multiplexing CRISPR/Cas9 in Saccharomyces cerevisiae Using Artificial Target DNA Sequences. Bio-protocol, 7(18).

Zhou J, et al. (2016) CasHRA (Cas9-facilitated Homologous Recombination Assembly) method of constructing megabase-sized DNA. Nucleic acids research, 44(14), e124.

Fu BX, et al. (2016) Distinct patterns of Cas9 mismatch tolerance in vitro and in vivo. Nucleic acids research, 44(11), 5365.