

# Resource Summary Report

Generated by [RRID](#) on Aug 19, 2026

## 3xFLAG-dCas9/pCMV-7.1

RRID:Addgene\_47948

Type: Plasmid

### Proper Citation

RRID:Addgene\_47948

### Plasmid Information

**URL:** <http://www.addgene.org/47948>

**Proper Citation:** RRID:Addgene\_47948

**Insert Name:** 3xFLAG-dCas9

**Bacterial Resistance:** Ampicillin

**Defining Citation:** [PMID:23942116](#)

**Vector Backbone Description:** Backbone Marker:Sigma-Aldrich; Backbone Size:4636; Vector Backbone:pCMV-7.1; Vector Types:Mammalian Expression, CRISPR; Bacterial Resistance:Ampicillin

**Comments:** This plasmid can be used to isolate specific genomic regions of interest using a catalytically inactive Cas9 fused with a tag(s). Purify: locus-specific chromatin immunoprecipitation (enChIP) This system is compatible with gRNA\_cloning\_vector ([www.addgene.org/41824](http://www.addgene.org/41824)) from the Church lab. Additional information and protocols can be found at: [http://www.med.hirosaki-u.ac.jp/~bgb/iChIP\\_protocols/index\\_e.html](http://www.med.hirosaki-u.ac.jp/~bgb/iChIP_protocols/index_e.html)

**Plasmid Name:** 3xFLAG-dCas9/pCMV-7.1

**Relevant Mutation:** human codon-optimized, D10A + H840A (catalytically inactive)

**Record Creation Time:** 20220422T222246+0000

**Record Last Update:** 20260815T081546+0000

### Ratings and Alerts

No rating or validation information has been found for 3xFLAG-dCas9/pCMV-7.1.

No alerts have been found for 3xFLAG-dCas9/pCMV-7.1.

## Data and Source Information

**Source:** [Addgene](#)

## Usage and Citation Metrics

We found 12 mentions in open access literature.

**Listed below are recent publications.** The full list is available at [RRID](#) .

Teague AG, et al. (2023) A polycistronic transgene design for combinatorial genetic perturbations from a single transcript in *Drosophila*. *PLoS genetics*, 19(6), e1010792.

Kwak Y, et al. (2022) Dynamic and widespread control of poly(A) tail length during macrophage activation. *RNA (New York, N.Y.)*, 28(7), 947.

Siadous FA, et al. (2021) Coxiella effector protein CvpF subverts RAB26-dependent autophagy to promote vacuole biogenesis and virulence. *Autophagy*, 17(3), 706.

Li K, et al. (2020) Interrogation of enhancer function by enhancer-targeting CRISPR epigenetic editing. *Nature communications*, 11(1), 485.

Huang Y, et al. (2020) Mass spectrometry-based proteomic capture of proteins bound to the MACC1 promoter in colon cancer. *Clinical & experimental metastasis*, 37(4), 477.

González-Rico FJ, et al. (2020) Alu retrotransposons modulate Nanog expression through dynamic changes in regional chromatin conformation via aryl hydrocarbon receptor. *Epigenetics & chromatin*, 13(1), 15.

Sher F, et al. (2019) Rational targeting of a NuRD subcomplex guided by comprehensive in situ mutagenesis. *Nature genetics*, 51(7), 1149.

Fujita T, et al. (2018) enChIP systems using different CRISPR orthologues and epitope tags. *BMC research notes*, 11(1), 154.

Hamidian A, et al. (2018) Promoter-associated proteins of EPAS1 identified by enChIP-MS - A putative role of HDX as a negative regulator. *Biochemical and biophysical research communications*, 499(2), 291.

Zuin J, et al. (2017) Regulation of the cohesin-loading factor NIPBL: Role of the lncRNA NIPBL-AS1 and identification of a distal enhancer element. *PLoS genetics*, 13(12), e1007137.

Fujii H, et al. (2015) Isolation of Specific Genomic Regions and Identification of Their Associated Molecules by Engineered DNA-Binding Molecule-Mediated Chromatin Immunoprecipitation (enChIP) Using the CRISPR System and TAL Proteins. *International journal of molecular sciences*, 16(9), 21802.

Fujita T, et al. (2015) Isolation of specific genomic regions and identification of associated molecules by engineered DNA-binding molecule-mediated chromatin immunoprecipitation (enChIP) using CRISPR. *Methods in molecular biology (Clifton, N.J.)*, 1288, 43.