

Resource Summary Report

Generated by [RRID](#) on Aug 20, 2026

pX603-AAV-CMV::NLS-dSaCas9(D10A,N580A)-NLS-3xHA-bGHpA

RRID:Addgene_61594

Type: Plasmid

Proper Citation

RRID:Addgene_61594

Plasmid Information

URL: <http://www.addgene.org/61594>

Proper Citation: RRID:Addgene_61594

Insert Name: hSaCas9

Organism: Other

Bacterial Resistance: Ampicillin

Defining Citation: [PMID:25830891](#)

Vector Backbone Description: Vector Backbone:pAAV; Vector Types:Mammalian Expression, AAV, CRISPR; Bacterial Resistance:Ampicillin

Comments: Note: This plasmid does not have an sgRNA expression cassette. Please refer to the associated publication, as well as the AAV portion of <http://www.addgene.org/crispr/zhang/> , for more information on expressing sgRNAs.

Plasmid Name: pX603-AAV-CMV::NLS-dSaCas9(D10A,N580A)-NLS-3xHA-bGHpA

Relevant Mutation: D10A, N580A

Record Creation Time: 20220422T222353+0000

Record Last Update: 20260815T081744+0000

Ratings and Alerts

No rating or validation information has been found for pX603-AAV-CMV::NLS-dSaCas9(D10A,N580A)-NLS-3xHA-bGHpA.

No alerts have been found for pX603-AAV-CMV::NLS-dSaCas9(D10A,N580A)-NLS-3xHA-bGHpA.

Data and Source Information

Source: [Addgene](#)

Usage and Citation Metrics

We found 16 mentions in open access literature.

Listed below are recent publications. The full list is available at [RRID](#) .

Kantor B, et al. (2024) The therapeutic implications of all-in-one AAV-delivered epigenome-editing platform in neurodegenerative disorders. Nature communications, 15(1), 7259.

Streb P, et al. (2023) How chromosomal translocations arise to cause cancer: Gene proximity, trans-splicing, and DNA end joining. iScience, 26(6), 106900.

Ouellette SP, et al. (2021) CRISPR Interference To Inducibly Repress Gene Expression in Chlamydia trachomatis. Infection and immunity, 89(7), e0010821.

Deutsch M, et al. (2021) AAV-Mediated CRISPRi and RNAi Based Gene Silencing in Mouse Hippocampal Neurons. Cells, 10(2).

Brockett MR, et al. (2021) A Dynamic, Ring-Forming Bactofilin Critical for Maintaining Cell Size in the Obligate Intracellular Bacterium Chlamydia trachomatis. Infection and immunity, 89(8), e0020321.

Nuñez JK, et al. (2021) Genome-wide programmable transcriptional memory by CRISPR-based epigenome editing. Cell, 184(9), 2503.

Præstholt SM, et al. (2021) Impaired glucocorticoid receptor expression in liver disrupts feeding-induced gene expression, glucose uptake, and glycogen storage. Cell reports, 37(5), 109938.

Wood NA, et al. (2020) The ClpX and ClpP2 Orthologs of Chlamydia trachomatis Perform Discrete and Essential Functions in Organism Growth and Development. mBio, 11(5).

Josipovi? G, et al. (2019) Active fusions of Cas9 orthologs. Journal of biotechnology, 301, 18.

Lau CH, et al. (2019) Targeted Transgene Activation in the Brain Tissue by Systemic Delivery of Engineered AAV1 Expressing CRISPRa. Molecular therapy. Nucleic acids, 16, 637.

Wurihan W, et al. (2019) Nonspecific toxicities of Streptococcus pyogenes and Staphylococcus aureus dCas9 in Chlamydia trachomatis. Pathogens and disease, 77(9).

Lu J, et al. (2018) Multimode drug inducible CRISPR/Cas9 devices for transcriptional activation and genome editing. *Nucleic acids research*, 46(5), e25.

Ouellette SP, et al. (2018) Feasibility of a Conditional Knockout System for Chlamydia Based on CRISPR Interference. *Frontiers in cellular and infection microbiology*, 8, 59.

Huang YA, et al. (2017) ApoE2, ApoE3, and ApoE4 Differentially Stimulate APP Transcription and A β Secretion. *Cell*, 168(3), 427.

Kleinjan DA, et al. (2017) Drug-tunable multidimensional synthetic gene control using inducible degron-tagged dCas9 effectors. *Nature communications*, 8(1), 1191.

Gao Y, et al. (2016) Complex transcriptional modulation with orthogonal and inducible dCas9 regulators. *Nature methods*, 13(12), 1043.