

Resource Summary Report

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pTF272 (pFA6a-TEV-6xGly-3xFlag-HphMX)

RRID:Addgene_44083

Type: Plasmid

Proper Citation

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Plasmid Information

URL: <http://www.addgene.org/44083>

Proper Citation: RRID:Addgene_44083

Bacterial Resistance: Ampicillin

Vector Backbone Description: Backbone Marker:Mark Hochstrasser lab (Yale Univeristy, New Haven, CT) (PMID: 19243080); Backbone Size:4477; Vector Backbone:pFA6a-6xGLY-3xFLAG-hphMX4 (Addgene plasmid # 20755); Vector Types:PCR-based yeast C-terminal tagging; Bacterial Resistance:Ampicillin

Comments: The depositing lab has constructed and deposited here 36 plasmids that can be used to generate linear PCR products to fuse to the 3' end of an ORF in the yeast genome and add (TEV or PreScission protease site) - 6X Gly linker - (12X His, 2X Strep, 3X FLAG, Protein A, or V5) - ADH1 transcription terminator - (KanMX, HphMX, or His3MX). Six plasmids (Addgene plasmids 44062, 44068, 44074, 44080, 44086, and 44092) have no epitope/purification tag and so can be used to add an alternate tag by standard cloning into unique XhoI-KpnI-XbaI sites, if desired. The other 30 plasmids are all of the possible combinations of the protease site - tag - marker options listed above. After fusion, the modified protein can then be followed with the epitope or purified using the tag and released from the matrix with the protease site. The constructs were designed with one of two different protease sites to allow a complex with two different tags to be purified in two successive steps, using each protease to release the complex from each purification matrix. The schematic diagram (see attached above) represents the format of the constructs and shows the proper scheme for producing the desired PCR products (see also below). If only epitope tagging is needed, the upstream primer can be adjusted to skip the protease site (see example below). Construction: DS5421 (pFA6a-6xGLY-3XFLAG-HIS3MX6 (Addgene Plasmid #20753), DS5422 (pFA6a-6xGLY-3XFLAG-KanMX) (Addgene Plasmid #20754), or DS5423 (pFA6a-6xGLY-3XFLAG-HphMX) (Addgene Plasmid #20755) (corresponding to each selection marker) was digested with AscI and PacI to release 3XFlag and replace it with TEV or PreScission sites followed by 6xGly and an XhoI-KpnI-XbaI linker. Affinity tags were constructed by PCR or by annealing oligos to

fit into the XhoI-XbaI gap. This includes 12xHis, 2xStrep II, 3xFLAG, PrtA (2 repeats as found in pBS1479), and V5. Each tag was inserted into each base vector, making 30 templates with all combinations of protease site, tag, and selectable marker. PCR amplification: These two primers (along with 50 nt of homology to the targets at the 5' ends) will amplify the desired product from any of the constructs (the reading frame for P1 is important – alternating codons bracketed). P1 5'-(homology)...[CGA]CGG[ATC]CCC[GGG]TTA[ATT]AAC-3' P2 5'-(homology)...GATATCATCGATGAATTCGAGCTCGTTT-3' If only epitope tagging is needed, the upstream primer can be adjusted to skip the protease site. As an example, for the V5-tag-containing constructs, usage of the following primer (5'-(homology)...GGTGGAGGCTCTAGAGGTAAGCCAATTCCA-3') in place of P1 will omit the protease cleavage site and add only 3xGly and V5 (GGGSR-V5-Stop).

Plasmid Name: pTF272 (pFA6a-TEV-6xGly-3xFlag-HphMX)

Record Creation Time: 20220422T222228+0000

Record Last Update: 20260815T081512+0000

Ratings and Alerts

No rating or validation information has been found for pTF272 (pFA6a-TEV-6xGly-3xFlag-HphMX).

No alerts have been found for pTF272 (pFA6a-TEV-6xGly-3xFlag-HphMX).

Data and Source Information

Source: [Addgene](#)

Usage and Citation Metrics

We found 3 mentions in open access literature.

Listed below are recent publications. The full list is available at [RRID](#).

Cooney I, et al. (2023) Lysate-to-grid: Rapid Isolation of Native Complexes from Budding Yeast for Cryo-EM Imaging. Bio-protocol, 13(2).

Christiano R, et al. (2020) A Systematic Protein Turnover Map for Decoding Protein Degradation. Cell reports, 33(6), 108378.

Farkas Z, et al. (2018) Hsp70-associated chaperones have a critical role in buffering protein production costs. eLife, 7.