Resource Summary Report

Generated by RRID on Aug 25, 2026

pY010 (pcDNA3.1-hAsCpf1)

RRID:Addgene_69982
Type: Plasmid

Proper Citation

RRID:Addgene_69982

Plasmid Information

URL: http://www.addgene.org/69982

Proper Citation: RRID:Addgene_69982

Insert Name: hAsCpf1

Organism: Acidaminococcus_sp_BV3L6

Bacterial Resistance: Ampicillin

Defining Citation: PMID:26422227

Vector Backbone Description: Backbone Size:5354; Vector Backbone:pcDNA3.1; Vector Types:Mammalian Expression, CRISPR; Bacterial Resistance:Ampicillin

Plasmid Name: pY010 (pcDNA3.1-hAsCpf1)

Record Creation Time: 20220422T222433+0000

Record Last Update: 20260815T081902+0000

Ratings and Alerts

No rating or validation information has been found for pY010 (pcDNA3.1-hAsCpf1).

No alerts have been found for pY010 (pcDNA3.1-hAsCpf1).

Data and Source Information

Source: Addgene

Usage and Citation Metrics

We found 61 mentions in open access literature.

Listed below are recent publications. The full list is available at RRID .

Kim S, et al. (2026) CRISPR-Cpf1-Mediated Gene-Editing System Based on a Single Bidirectional Promoter. International journal of molecular sciences, 27(10).

Nikitchina N, et al. (2025) Targeted deletions in human mitochondrial DNA engineered by Type V CRISPR-Cas12a system. NAR molecular medicine, 2(2), ugaf021.

Chen P, et al. (2025) Highly parallel profiling of the activities and specificities of Cas12a variants in human cells. Nature communications, 16(1), 3022.

Raghavan R, et al. (2025) Rational engineering of minimally immunogenic nucleases for gene therapy. Nature communications, 16(1), 105.

Chen P, et al. (2024) Engineering of Cas12a nuclease variants with enhanced genome-editing specificity. PLoS biology, 22(3), e3002514.

Wang H, et al. (2024) Engineering of a compact, high-fidelity EbCas12a variant that can be packaged with its crRNA into an all-in-one AAV vector delivery system. PLoS biology, 22(5), e3002619.

Slaman E, et al. (2024) Comparison of Cas12a and Cas9-mediated mutagenesis in tomato cells. Scientific reports, 14(1), 4508.

Illa-Berenguer E, et al. (2023) Editing efficiencies with Cas9 orthologs, Cas12a endonucleases, and temperature in rice. Frontiers in genome editing, 5, 1074641.

Zhou J, et al. (2022) Cas12a variants designed for lower genome-wide off-target effect through stringent PAM recognition. Molecular therapy : the journal of the American Society of Gene Therapy, 30(1), 244.

Ou T, et al. (2022) Reprogramming of the heavy-chain CDR3 regions of a human antibody repertoire. Molecular therapy : the journal of the American Society of Gene Therapy, 30(1), 184.

Blomme J, et al. (2022) The heat is on: a simple method to increase genome editing efficiency in plants. BMC plant biology, 22(1), 142.

Shin HR, et al. (2021) Gene Manipulation Using Fusion Guide RNAs for Cas9 and Cas12a. Methods in molecular biology (Clifton, N.J.), 2162, 185.

Duan N, et al. (2021) An Episomal CRISPR/Cas12a System for Mediating Efficient Gene Editing. Life (Basel, Switzerland), 11(11).

Wang X, et al. (2021) A far-red light-inducible CRISPR-Cas12a platform for remote-controlled genome editing and gene activation. Science advances, 7(50), eabh2358.

Berkhout B, et al. (2021) Design and Evaluation of Guide RNA Transcripts with a 3'-Terminal HDV Ribozyme to Enhance CRISPR-Based Gene Inactivation. Methods in molecular biology (Clifton, N.J.), 2167, 205.

McMahon MA, et al. (2021) Gene Disruption Using Chemically Modified CRISPR-Cpf1 RNA. Methods in molecular biology (Clifton, N.J.), 2162, 49.

Ageely EA, et al. (2021) Gene editing with CRISPR-Cas12a guides possessing ribose-modified pseudoknot handles. Nature communications, 12(1), 6591.

Sherstyuk VV, et al. (2021) Generation of Transgenic Rat Embryonic Stem Cells Using the CRISPR/Cpf1 System for Inducible Gene Knockout. Biochemistry. Biokhimiia, 86(7), 843.

Tran MH, et al. (2021) A more efficient CRISPR-Cas12a variant derived from Lachnospiraceae bacterium MA2020. Molecular therapy. Nucleic acids, 24, 40.

Chen P, et al. (2020) A Cas12a ortholog with stringent PAM recognition followed by low off-target editing rates for genome editing. Genome biology, 21(1), 78.