

Resource Summary Report

Generated by [RRID](#) on Sep 10, 2026

Forsyth Institute Advanced Microscopy Core Facility

RRID:SCR_021121

Type: Tool

Proper Citation

Forsyth Institute Advanced Microscopy Core Facility (RRID:SCR_021121)

Resource Information

URL: <https://imaging.forsyth.org/>

Proper Citation: Forsyth Institute Advanced Microscopy Core Facility (RRID:SCR_021121)

Description: Offers microscopy imaging services to both internal and external users in the greater Boston area. The facility is equipped for live cell imaging, laser scanning confocal and super resolution microscopy. Common sample types imaged in the facility are live and fixed bacteria, biofilms, embedded soft tissue, cleared tissue, and mineralized tissue and samples can be labelled or label free. One area of expertise is imaging many fluorophores simultaneously using spectral imaging and linear unmixing. In addition to instrument training, we can also provide advice for sample preparation and image analysis. In some cases, we are able to provide full-service imaging for customers.

Synonyms: Forsyth Institute Imaging Core, The Forsyth Institute Forsyth-Advanced Microscopy Core, Forsyth-Advanced Microscopy Core

Resource Type: access service resource, core facility, service resource

Keywords: USEdit, microscopy imaging services, live cell imaging, laser, scanning, confocal, super resolution microscopy, ABRF, ABRF

Availability: open

Resource Name: Forsyth Institute Advanced Microscopy Core Facility

Resource ID: SCR_021121

Alternate IDs: ABRF_1159

Alternate URLs: <https://coremarketplace.org/?FacilityID=1159>

Record Creation Time: 20220129T080353+0000

Record Last Update: 20260905T133420+0000

Ratings and Alerts

No rating or validation information has been found for Forsyth Institute Advanced Microscopy Core Facility.

No alerts have been found for Forsyth Institute Advanced Microscopy Core Facility.

Data and Source Information

Source: [SciCrunch Registry](#)

Usage and Citation Metrics

We found 9 mentions in open access literature.

Listed below are recent publications. The full list is available at [RRID](#) .

Nahar N, et al. (2026) Ultrasmall episymbiont Nanosynbacter lyticus employs multiple ATP-generating metabolic pathways during horizontal transmission. The ISME journal, 20(1).

Gundrum J, et al. (2026) Slicing overcomes the bacterial cell wall barrier to fluorescence in situ hybridization. Microbiology spectrum, 14(1), e0200125.

Martín A, et al. (2026) Targeted KRASG12V Degradation in vivo Elicits Lung Adenocarcinoma Regression with Subsequent Relapse from Dysregulated Proteolysis. Cancer research.

Grossman AS, et al. (2024) Saccharibacteria deploy two distinct Type IV pili, driving episymbiosis, host competition, and twitching motility. bioRxiv : the preprint server for biology.

Ramirez-Puebla ST, et al. (2024) Improved Visualization of Oral Microbial Consortia. Journal of dental research, 103(13), 1421.

Depalle B, et al. (2023) Rapid post-eruptive maturation of porcine enamel. Frontiers in physiology, 14, 1099645.

Gil-Bona A, et al. (2023) Proteomic Analyses Discern the Developmental Inclusion of Albumin in Pig Enamel: A New Model for Human Enamel Hypomineralization. International journal of molecular sciences, 24(21).

Matute JD, et al. (2023) Intelectin-1 binds and alters the localization of the mucus barrier-modifying bacterium Akkermansia muciniphila. The Journal of experimental medicine, 220(1).

Chipashvili O, et al. (2021) Episymbiotic Saccharibacteria suppresses gingival inflammation and bone loss in mice through host bacterial modulation. Cell host & microbe, 29(11), 1649.