

Resource Summary Report

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University of Pennsylvania Perelman School of Medicine Cytomics and Cell Sorting Resource Laboratory Core Facility

RRID:SCR_022376

Type: Tool

Proper Citation

University of Pennsylvania Perelman School of Medicine Cytomics and Cell Sorting Resource Laboratory Core Facility (RRID:SCR_022376)

Resource Information

URL: <https://pathbio.med.upenn.edu/pbr/portal/flowcyto/>

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Description: Flow cytometry shared resource laboratory at the University of Pennsylvania. Facility has instruments, which include analyzers, cell sorters, small particle detectors, dual fluorescence cell counter/viability instrument, tissue dissociator for cell preparation. Provides on-site and off-site support to instrument users, including analyzer and cell sorter training. Core's Research and Development team collaborates/consults with principal investigators in developing high-dimensional panels, as well as staining, acquisition, and analysis.

Synonyms: Penn Cytomics, Cytomics and Cell Sorting Resource Laboratory, University of Pennsylvania Perelman School of Medicine Cytomics and Cell Sorting Resource Laboratory

Resource Type: access service resource, core facility, service resource

Keywords: ABRF, flow cytometry, cell sorting, analyzer and cell sorter training,

Funding: NCI P30 016520

Availability: Open

Resource Name: University of Pennsylvania Perelman School of Medicine Cytomics and Cell Sorting Resource Laboratory Core Facility

Resource ID: SCR_022376

Alternate IDs: ARBF_1389

Alternate URLs: <https://coremarketplace.org?citation=1&FacilityID=1389>

Record Creation Time: 20220602T050140+0000

Record Last Update: 20260905T133425+0000

Ratings and Alerts

No rating or validation information has been found for University of Pennsylvania Perelman School of Medicine Cytomics and Cell Sorting Resource Laboratory Core Facility.

No alerts have been found for University of Pennsylvania Perelman School of Medicine Cytomics and Cell Sorting Resource Laboratory Core Facility.

Data and Source Information

Source: [SciCrunch Registry](#)

Usage and Citation Metrics

We found 79 mentions in open access literature.

Listed below are recent publications. The full list is available at [RRID](#) .

Merolle M, et al. (2026) The role of integrins in T cell-mediated resistance to *Cryptosporidium parvum*. *bioRxiv* : the preprint server for biology.

Maslanka JR, et al. (2026) Generation and characterization of a novel MHC-II tetramer for tracking and characterization of toxin B-specific CD4+ T cell responses. *bioRxiv* : the preprint server for biology.

Liang B, et al. (2026) Early clonal dominance at priming sets the trajectory for broad HIV serum neutralization. *bioRxiv* : the preprint server for biology.

Hamilton AG, et al. (2026) High-Throughput In Vivo Screening Using Barcoded mRNA Identifies Lipid Nanoparticles With Extrahepatic Tropism for In Situ Immunoengineering. *Advanced materials* (Deerfield Beach, Fla.), 38(13), e14370.

Marzolini N, et al. (2026) Targeting DNA-LNPs to Endothelial Cells Improves Expression Magnitude, Duration, and Specificity. *Advanced science* (Weinheim, Baden-Wurttemberg, Germany), 13(14), e15480.

Mukalel AJ, et al. (2026) Bioinspired oxidized mRNA lipid nanoparticles for ex vivo engineering of chimeric antigen receptor macrophages targeting solid tumors. *Bioengineering & translational medicine*, 11(3), e70138.

Murthy A, et al. (2026) Regulatory T cells protect against aberrant remodeling in a mouse model of pulmonary fibrosis. *Mucosal immunology*, 19(2), 1769.

Wu L, et al. (2026) ImmunoPheno: A Computational Framework for Data-Driven Design and Analysis of Immunophenotyping Experiments. *bioRxiv : the preprint server for biology*.

Geisler HC, et al. (2026) Resolving heterogeneity of targeted lipid nanoparticles through solution-based biophysical analyses. *bioRxiv : the preprint server for biology*.

Brysgel TV, et al. (2026) Decoupling Physisorption from Chemisorption in Clickable Lipid Nanoparticles. *ACS nanoscience Au*, 6(3), 348.

Zeng J, et al. (2026) Rapid receptor internalization potentiates CD7-targeted lipid nanoparticles for efficient mRNA delivery to T cells and in vivo CAR T-cell engineering. *bioRxiv : the preprint server for biology*.

Vrathasha V, et al. (2026) Variant-to-gene mapping identifies ARHGEF12 as a primary open-angle glaucoma effector gene operating within retinal ganglion cells. *bioRxiv : the preprint server for biology*.

Deng MZ, et al. (2026) Cell-derived Nanoparticles Provide a Robust Platform to Manufacture Therapeutic T cells. *Research square*.

Foos KM, et al. (2026) Generation of functional canine TIL products for solid tumors. *Frontiers in immunology*, 17, 1810955.

Frazee CS, et al. (2026) Fate induction through asymmetric T cell division is modulated by chimeric antigen receptor costimulatory domains. *Nature immunology*, 27(7), 1433.

Espy CL, et al. (2026) Localized NF- κ B Inhibition Reduces Lipid Nanoparticle-Associated Inflammation. *Advanced science (Weinheim, Baden-Wurttemberg, Germany)*, 13(31), e17931.

Kim D, et al. (2026) Crosslinked ionizable lipids reprogram dendritic cell metabolism for potent mRNA vaccination. *Nature materials*.

Geisler HC, et al. (2026) Resolving Heterogeneity of Targeted Lipid Nanoparticles Through Solution-Based Biophysical Analyses. *Advanced materials (Deerfield Beach, Fla.)*, e73984.

Maltez VI, et al. (2026) Agonistic anti-CD40 antibody treatment converts resident regulatory T cells into activated type 1 effectors within the tumor microenvironment. *Immunity*, 59(4), 1058.

Barnes DT, et al. (2026) Delta family protocadherins contribute to protoglomerular targeting of olfactory sensory neuron axons in the olfactory bulb. *PLoS genetics*, 22(4), e1012090.