

Resource Summary Report

Generated by RRID on Aug 5, 2026

CRISPResso2

RRID:SCR_024503

Type: Tool

Proper Citation

CRISPResso2 (RRID:SCR_024503)

Resource Information

URL: <https://github.com/pinellolab/CRISPResso2>

Proper Citation: CRISPResso2 (RRID:SCR_024503)

Description: Software pipeline designed to enable rapid and intuitive interpretation of genome editing experiments. Used for analysis of deep sequencing data for rapid and intuitive interpretation of genome editing experiments.

Resource Type: data processing software, software application, software resource, data analysis software

Keywords: sequencing data analysis, genome editing experiments interpretation,

Availability: Free, Available for download, Freely available

Resource Name: CRISPResso2

Resource ID: SCR_024503

License URLs: <https://github.com/pinellolab/CRISPResso2/blob/master/LICENSE.txt>

Record Creation Time: 20231002T161336+0000

Record Last Update: 20260805T044532+0000

Ratings and Alerts

No rating or validation information has been found for CRISPResso2.

No alerts have been found for CRISPResso2.

Data and Source Information

Usage and Citation Metrics

We found 44 mentions in open access literature.

Listed below are recent publications. The full list is available at [RRID](#) .

Raguram A, et al. (2025) Directed evolution of engineered virus-like particles with improved production and transduction efficiencies. *Nature biotechnology*, 43(10), 1635.

Wang Y, et al. (2025) CRISPR-Enabled Autonomous Transposable Element (CREATE) for RNA-based gene editing and delivery. *EMBO reports*, 26(4), 1062.

Liao K, et al. (2025) exoCasMINI: A T5 exonuclease fused CRISPR-Cas12f system with enhanced gene editing efficiency. *iScience*, 28(8), 113171.

Shirole NH, et al. (2025) Requirement for Cyclin D1 Underlies Cell-Autonomous HIF2 Dependence in Kidney Cancer. *Cancer discovery*, 15(7), 1484.

Pandey S, et al. (2025) Efficient site-specific integration of large genes in mammalian cells via continuously evolved recombinases and prime editing. *Nature biomedical engineering*, 9(1), 22.

Chen K, et al. (2025) Lung and liver editing by lipid nanoparticle delivery of a stable CRISPR-Cas9 ribonucleoprotein. *Nature biotechnology*, 43(9), 1445.

Shi H, et al. (2025) Rapid two-step target capture ensures efficient CRISPR-Cas9-guided genome editing. *Molecular cell*, 85(9), 1730.

Sousa AA, et al. (2025) Systematic optimization of prime editing for the efficient functional correction of CFTR F508del in human airway epithelial cells. *Nature biomedical engineering*, 9(1), 7.

Durbacz MZ, et al. (2025) Optimized genomic editing of a common Duchenne muscular dystrophy mutation in patient-derived muscle cells and a new humanized mouse model. *Molecular therapy. Nucleic acids*, 36(2), 102569.

Netsawang C, et al. (2025) Precise correction of G6PD Viangchan mutation in iPSCs by prime editing strategy. *Scientific reports*, 15(1), 30192.

Song Z, et al. (2025) Noncanonical target-strand cytosine base editing via engineered Un1Cas12f1 platform. *Nature communications*, 16(1), 9499.

Pierce SE, et al. (2025) Prime editing-installed suppressor tRNAs for disease-agnostic genome editing. *Nature*, 648(8092), 191.

Kiseleva AA, et al. (2025) Fine tuning wheat heading time through genome editing of transcription factor binding sites in Ppd-1 gene promoter. *Scientific reports*, 15(1), 42034.

Lee S, et al. (2025) Combining Multiplexed CRISPR/Cas9-Nickase and PARP Inhibitors Efficiently and Precisely Targets Cancer Cells. *Cancer research*, 85(15), 2890.

Takahashi G, et al. (2025) High-throughput robotic isolation of human iPS cell clones reveals frequent homozygous induction of identical genetic manipulations by CRISPR-Cas9. *Stem cell research & therapy*, 16(1), 295.

Ling S, et al. (2025) Customizable virus-like particles deliver CRISPR-Cas9 ribonucleoprotein for effective ocular neovascular and Huntington's disease gene therapy. *Nature nanotechnology*, 20(4), 543.

Meng R, et al. (2025) Engineered Cas12j-8 is a Versatile Platform for Multiplexed Genome Modulation in Mammalian Cells. *Advanced science* (Weinheim, Baden-Wurttemberg, Germany), 12(33), e02593.

Herger M, et al. (2025) High-throughput screening of human genetic variants by pooled prime editing. *Cell genomics*, 5(4), 100814.

du Rand A, et al. (2025) Efficient Dual Cas9 Nickase Correction of a Prevalent Pathogenic LAMB 3 Variant for Junctional Epidermolysis Bullosa. *JID innovations : skin science from molecules to population health*, 5(3), 100343.

Gowen BG, et al. (2025) Systematic identification and characterization of high efficiency Cas9 guide RNAs for therapeutic targeting of ADAR. *PloS one*, 20(2), e0317745.